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Optogeny of Adrenergic Arousal and Cholinergic Inhibitory Mechanisms in the Rat

Abstract. With spontaneous activity as a measure of arousal, dose response curves were established for scopolamine and amphetamine administered to 10-, 15-, 20-, 25-, and 100-day-old rats. Amphetamine always increased activity, but scopolamine had no effect on younger rats, which suggests that adrenergic excitatory areas in the brainstem mature more rapidly than cholinergic inhibitory areas in the forebrain.

Generalized excitatory and inhibitory systems in the brain regulate overall levels of arousal. The major excitatory center is thought to be the brainstem reticular formation. When activity in this area is destroyed by lesions, stupor results (1); when the area is activated by electrical stimulation, electrocortical and behavioral arousal occurs (2). Acting in opposition to this excitatory region are certain forebrain structures which serve to modulate reticular excitability. When these centers or their connections with the brainstem are impaired, the effects of reticular stimulants are greatly augmented (3). When this area is activated by electrical stimulation, arousal is depressed (4).

The biochemical substrates of the arousal areas in the hindbrain and inhibitory centers in the forebrain are distinct, with the former primarily adrenergic in nature and the latter predominantly cholinergic (5). Amphetamine, which mimics adrenergic transmission by release of norepinephrine (6), particularly in the brainstem (7). induces large increments in locomotor activity, while adrenolytic agents depress arousal and generally lead to sedation (8). The anticholinergic drug scopolamine, which blocks acetylcholine transmission by occupation of postsynaptic sites (9), produces marked increments in activity (10), while anticholinesterases (11) and cholinomimetic agents (12) depress arousal.

It is now generally accepted that the development of the brain proceeds rostrally with phylogenetically primitive hindbrain structures maturing earlier than the younger forebrain systems (13). Thus neonatal animals should pass through a phase during which they are responsive to reticular stimulants and unaffected by cholinergic blocking agents because of the functional absence of forebrain inhibitory mechanisms. We now show that the neonatal rat is responsive to the reticular stimulant amphetamine, before it is responsive to scopolamine, an innibitor of forebrain cholinergic activity.

Degree of behavioral arousal was measured in stabilimeter activity cages scaled to the size of the animal. The largest cages, those used for the adults (14) consisted of wire mesh cages, 17.5 by 20.0 by 37.5 cm, mounted on a central axle which permitted the cage to tip slightly and activate a sensitive switch as the rat moved from one end to the other. For 10-day-old rats the cage dimensions were scaled down to 6.3 by 7.5 by 13.7 cm, and for 15-, 20-, and 25-day-old rats the cage was 8.7 by 10.0 by 18.7 cm. The activity cages were housed in temperaturecontrolled cubicles maintained at 29°C for the 10-, 15-, 20-, and 25-day-old rats and at 22°C for the 100-day-old

Dose response curves for both aniphetamine and scopolamine were determined at five different ages: 10, 15, 20, 25, and 100 days. Atreach describ 10-764 http://createstail.com/cookeratemins tested renly concerned total of 772 Sprague-Dawley rats were used, half of which were male and half female.

Rats were removed from living cages in a central colony room, placed in the activity cages for a 30-minute habituation period, and then injected with onedose of either d-amphetamine sulfate (0.250, 0.50, 1.0, 2.0, 4.0, or 8.0 mg/kg, salt weight), scopolamine hydrochloride (0.125, 0.250, 0.50, 1.0, 2.0, or 4.0 mg/kg, salt weight) or an equivalent volume of the 0.9 percent saline vehicle. They were then returned to the cages for 2 hours; during this time the number of crossings was recorded on printing counters every half hour. In addition, methylscopolamine hydrobromide (0.125, 0.250, 0.50, 1.0, 2.0 or 4.0 mg/kg, adjusted for equivwas administered to a group of 25-day- sponse in the older animals. old rats for control of the possible per kilogram of body weight.

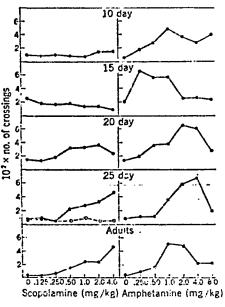


Fig. 1. The effects of scopolamine hydrochloride and d-amphetamine sulfate on spontaneous activity of rats of five different ages. The effect of methyl scopolamine is shown by the dotted line in the 25-day

activity occurring during the entire 2hour test period for all groups. Amphetamine produced an increment in activity, proportional to desage, in animals at all of the ages studied, while scopolamine increased activity only in animais 20 days of age and older. Methylscopolamine had no systematic effect on activity, which indicates that the scopolamine-induced increase in activity was the result of central rather than peripheral effects. No consistent sex differences in response to the drugs were found except in the 100-day-old group, where the females were more active regardless of whether drugs were given or not.

The results also suggest that the maximum effective dose of amphetamine was somewhat dependent on age. The 15-day group appeared to be more sensitive to amphetamine than the older animals insofar as they showed maxialent amounts of scopolamine), a drug mum activity increases in response to which does not cross the blood-brain low dosages, whereas higher dosages barrier in significant quantities (15), were required to elicit maximum re-

These results demonstrate that reacperipheral effects produced by the sco-tivity to amphetamine and scopolamine polamine hydrochloride. All drugs and matures at different rates in the neothe saline control were administered natal rat. To interpret these results, it intraperitoneally in a volume of 1 ml is most plausible to assume that amphetamine acts by releasing endogenous Figure 1 shows the mean amount of norepinephrine in primitive hindbrain

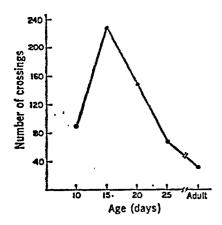


Fig. 2. Spontaneous activity as a function of age in nondrugged animals.

arousal centers and that these centers mature earlier than the forebrain inhibitory centers do. In turn, scopolamine cannot act to block the cholinergic forebrain inhibitory centers until these centers are functionally mature and exerting a chronic inhibitory influence on hindbrain activity.

Additional evidence for the delayed maturation of forebrain inhibitory centers is found in the activity pattern of the saline control groups. When saline control groups of the same age) novel environment reaches a peak in hibitory mechanism is cholinergic. rats at about 15 days of age and declines rapidly in the subsequent 10 days. The increase in activity between days 10 and 15 undoubtedly reflects in- Department of Psychology, creasing skeletal muscular development Princeton University, plus increasing sensory responsiveness Princeton, New Jersey 08540 (16). The decline in normal activity corresponds strikingly to the increasing effectiveness of scopolamine as a be- 1. D. B. Lindsley, J. Bowden, H. W. Magoun,

haviorally arousing drug, which suggests that forebrain cholinergic inhibitory centers also act to modulate exploratory activity in novel environ-

Moreover, this period of declining arousal and increasing sensitivity to anticholinergies also parallels functional development of the forebrain. Primitive electroencephalographic activity is first noted at 6 days after birth, but the spectral composition does not appreximate that of the adult until the rat is between 25 and 30 days of age (17). Myelin, which is present in the brainstem at birth, is not seen in the forebrain until 10 days after birth, with the greatest deposition occurring between 15 and 30 days of age (18). Similarly, the number of synaptic junctions in the cortex undergoes massive proliferation between 15 and 25 days of age (19).

Considerable evidence from studies of humans supports the view that the forebrain areas exert inhibitory control over hindbrain mechanisms (26). Both the human infant and the rat display little more than simple involuntary responses at birth. With maturation, many of these reflexes disappear and then rethese data (combined means for all appear with corticol atrophy in senescence or after cortical injury (21). are plotted separately so that the age- Forebrain development thus appears to related trends are not masked by the modulate both primitive reflexes and larger drug effects (Fig. 2), it is ap- behavioral arousal. Our data suggest parent that spontaneous activity in a that at least some portion of this in-

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References and Notes

Electroencephalogr. Clin. Neurophysiol. 1, 475 (1949).

2. G. Moruzzi and H. W. Magoun, ibid. p. 455. A. Hugelin, M. Bonvallet, P. Dell, ibid. 11, 325 (1989); G. S. Lynch, P. Ballantine, W. Levy, B. 195 (1969). A. Campbell, Exp. Neurol. 23,

4. P. Dell, in Brain Mechanisms, G. Moruzzi, A. Fessard, H. H. Jasper, Eds. (Elsevier, New York, 1963); C. D. Clemente, Cond. Reflex 3, 145 (1968).

5. Evidence for the adrenergic basis of arousal stems from many experimental techniques and is widely accepted [P. B. Bradley and J. Elkes, Brain 80, 77 (1957); J. J. Schildkraut and S. S. Kety, Science 156, 21 (1967)], although 50, universally [A. J. Mandell and C. E. Spooner, thid. 162, 142 (1968). In contrast, the view that the inhibitory functions of the forebrain are mediated by cholinergic transmission has not been ex-plicitly stated, although ample evidence for this view exists. For example, neuro-pharmacological studies have shown (1) high concentrations of acetylcholine in the fore-brain [H. McLennan, Synaptic Transmission (Saunders, Philadelphia, 1963), pp. 69-76]; (ii) cholinoceptive cell bodies in the cortex [K. Krnjevic and J. W. Phillis, J. Physiol. 328 (1963)]; (iii) increased release of acetylcholine in the cortex during arousal IT. Kanai and J. C. Szerb, Nature 205, 80 (1965); J. W. Phillis, Brain Res. 7, 378 (1968)]; and (iv) innervation of many fore-brain structures by cholinergic fibers crigibrain structures by cholinercic fibers originating in the brainstem [C. C. D. Shute and P. R. Lewis, Brain 90, 497 (1967)].

6. L. C. F. Hanson, Psychopharmacologia 10, 289 (1967).

7. F. Javoy, A. M. Thierry, S. S. Kety, J. Glowinski, Commun. Behav. Biol. 1, 43 (1968).

8. For example, L. C. F. Hanson, Psychopharmacologia 8, 100 (1965).

9. N. J. Giarman and G. Pepeu, Brit. J. Phalmacol. Chemother. 23, 123 (1964).

10. N. Pradhan and T. Roth, Psychopharmacologia 12, 358 (1968).

S. N. Pradhan and T. Roth, Psychopharmacologia 12, 358 (1968).
 W. R. Price and S. F. Dorbel, Psychon. Sci 8, 117 (1967).

12. G. Zetler, Int. J. Neuropharmacol. 7, 325 (1968) 13. D. Richter, in Regional Development of the Brain in Early Life, A. Minkowski, Ed. (Davis, Philadelphia, 1957).

14. B. A. Campbell, in Thirst, M. Wayner, Ed. (Pergamon, New York, 1964).

15. J. Goodman and A. Gilman, Ed. The

15. L. S. Goodman and A. Gilman, Eds., The Pharmacological Busis of Therapeutics (Macmillan, New York, ed. 3, 1965), p. 336.

R. C. Bolles and P. J. Woods, Anim. Behav. 12, 427 (1964).

L. Deza and E. Eidelberg, Exp. Neurol. 17, 425 (1967).
 S. Jacobson, J. Comp. Neurol. 121, 5 (1963).

19. G. K. Agajuhanian and F. E. Bloom, Brain Res. 6, 716 (1967).

20. G. Bronson, Behav. Sci. 10, 7 (1905)

 D. Denny-Blown, J. Nerv. Ment. Dis. 126, 9 (1988); G. Paulson and G. Gottlieb. Brain 91, 37 (1968).

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